

4-Substituted cyclohexyl sulfones as potent, orally active γ -secretase inhibitors

Ian Churcher,^{a,*} Dirk Beher,^b Jonathan D. Best,^c José L. Castro,^a Earl E. Clarke,^b Amy Gentry,^a Timothy Harrison,^a Laure Hitzel,^a Euan Kay,^a Sonia Kerrad,^a Huw D. Lewis,^b Pablo Morentin-Gutierrez,^a Russell Mortishire-Smith,^a Paul J. Oakley,^a Michael Reilly,^a Duncan E. Shaw,^a Mark S. Shearman,^b Martin R. Teall,^a Susie Williams^a and Jonathan D. J. Wrigley^b

^aDepartment of Medicinal Chemistry, The Neuroscience Research Centre, Merck Sharp and Dohme Research Laboratories, Terlings Park, Eastwick Road, Harlow, Essex CM20 2QR, UK

^bDepartment of Molecular and Cellular Neuroscience, The Neuroscience Research Centre, Merck Sharp and Dohme Research Laboratories, Terlings Park, Eastwick Road, Harlow, Essex CM20 2QR, UK

^cDepartment of In Vivo Neuroscience, The Neuroscience Research Centre, Merck Sharp and Dohme Research Laboratories, Terlings Park, Eastwick Road, Harlow, Essex CM20 2QR, UK

Received 28 September 2005; revised 3 October 2005; accepted 4 October 2005
Available online 3 November 2005

Abstract—The protease γ -secretase plays a pivotal role in the synthesis of pathogenic amyloid- β in Alzheimer's disease. Here, we report a further extension to a series of cyclohexyl sulfone-based γ -secretase inhibitors which has allowed the preparation of highly potent compounds which also demonstrate robust A β (40) lowering in vivo (e.g., compound **32**, MED 1 mg/kg p.o. in APP-YAC mice).

© 2005 Elsevier Ltd. All rights reserved.

Alzheimer's disease (AD) is a devastating disorder of the aged population and with current palliative treatments being of modest efficacy, the need remains for a therapy able to alter the underlying pathophysiology of the disease. The predominantly 40–42 amino acid amyloid- β (A β) peptide is the major component of the extracellular proteinaceous plaques seen in Alzheimer's disease (AD) and much evidence suggests a pivotal role for A β in the disease process.¹ In particular, individuals possessing autosomal dominant mutations in the genes encoding for amyloid- β precursor protein (β APP) or the membrane-bound protein homologs presenilin 1 and 2 have elevated A β levels and suffer from aggressive forms of early onset AD.^{2,3} These observations have led to the hypothesis that A β , either in its soluble form or when

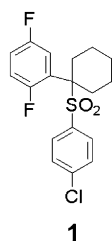
aggregated into oligomers, fibrils, and subsequently plaques, is responsible for neuronal toxicity and cell death.⁴

A β is derived by processing of the 695–770 residue, type I transmembrane protein β APP.⁵ The major metabolic pathway of β APP involves sequential cleavage by the proteases α -secretase and γ -secretase leading to non-amyloidogenic fragments. Alternative processing by stepwise cleavage mediated by β -secretase and γ -secretase leads to the production of A β and it is inhibitors of the latter enzyme that were targeted in the current work.^{6,7}

γ -Secretase is a novel membrane-bound aspartyl protease complex formed from presenilin-1/2, nicastrin, Aph-1, and pen-2 whose catalytic center is unusual in likely being within transmembrane helices of the presenilin unit. As the role of γ -secretase has become better characterized, a variety of substrates⁸ including the Notch receptor have been identified leading to the possibility of mechanism-based effects⁹ or alternatively to additional therapeutic uses¹⁰ for γ -secretase inhibitors.

Keywords: Alzheimer's disease; γ -Secretase; Protease inhibitor; Amyloid; In vivo.

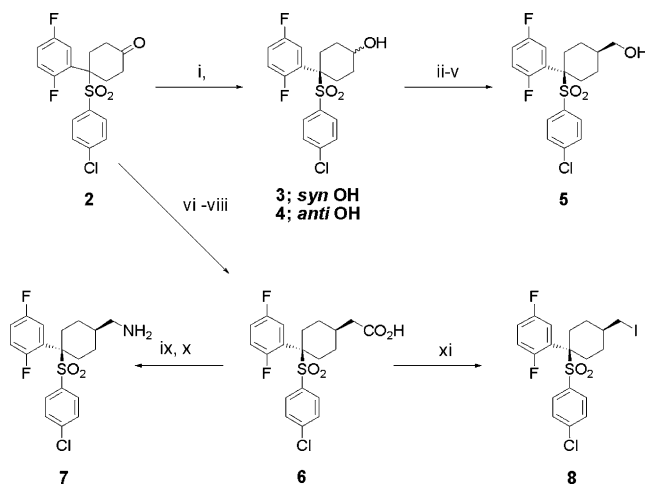
* Corresponding author. Tel.: +44 1279 440000; fax: +44 1279 440390; e-mail: ian_churcher@merck.com



A whole-cell γ -secretase inhibition assay using SH-SY5Y neuroblastoma cells in which human γ -secretase catalyzes the breakdown of the overexpressed exogenous substrate A4CTF has been developed.¹¹ We have previously disclosed^{12–15} a series of cyclohexyl sulfones which have demonstrated *in vitro* inhibition of A β (40) secretion in this assay. In this letter, we describe the further optimization of these leads resulting in the identification of highly potent analogs demonstrating *in vivo* reduction of brain A β (40) in a mouse model.

Based upon the 1,1-disubstituted cyclohexane **1**, which has previously been shown to be a potent γ -secretase inhibitor (IC₅₀ 3 nM),¹² we chose to probe the 4-position on the cyclohexane ring in order to improve both potency at γ -secretase and also ADME properties.

Investigation of substitution at the 4-position was facilitated by the ready availability, on scale, of the cyclohexanone derivative **2**.¹² This could be elaborated to provide a range of useful intermediates as shown in Scheme 1. Reduction of **2** gave predominantly the *anti*-alcohol **4** which underwent smooth cyanide displacement via a mesylate to give, after hydrolysis and reduction, *syn* hydroxymethyl **5**. Alternatively, Wadsworth–Horner–Emmons reaction followed by stereoselective reduction and hydrolysis gave the carboxylate **6**, which was elaborated to amine **7** via Curtius rearrangement or iodide **8** by Hunsdiecker-type radical iodination.¹⁶



Scheme 1. Reagents and conditions: (i) NaBH₄, EtOH, 10 °C (anti:syn 6:1); (ii) MsCl, Et₃N, DCM, –50 °C; (iii) Bu₄CN, PhMe, 75 °C; (iv) AcOH/concd HCl, 110 °C; (v) ^tBuOCOCl, Et₃N, THF then NaBH₄, H₂O; (vi) (EtO)₂P(O)CH₂CO₂Et, NaH, THF; (vii) L-Selectride, THF, –40 °C; (viii) LiOH, aq MeOH; (ix) (COCl)₂, DMF, THF then NaN₃, Bu₄NBr, PhH then BnOH, reflux; (x) HBr/AcOH then NaOH; (xi) PhI(OAc)₂, I₂, PhH, reflux, hv.

range or iodide **8** by Hunsdiecker-type radical iodination.¹⁶

These intermediates could be further elaborated to a wide range of derivatives, a representative number of which are summarized in Table 1. Whilst the hydroxymethyl analog **5** showed activity similar to that of unsubstituted **1** only, formation of a carbamate (e.g., compound **9**) gave an increase in potency. Similarly, the poor activity of primary amine **7** could be improved markedly by acylation or sulfonylation with ureas, carbamates, and sulfamides all being well tolerated. Additionally, several heterocycles including succinimide provided compounds with good activity.

The iodide **8** also allowed ready access to either sulfonamides (via KNO₃/SO₂Cl₂ oxidation¹⁷ of the intermediate thiol) or sulfones (via S_N2 displacement of the iodide and subsequent oxidation) as shown in Scheme 2. The *in vitro* potencies of a representative range of sulfonyl-containing analogs are summarized in Table 2.

The series of simple alkyl and aryl sulfones gave mostly modest inhibitory activity (data not shown) although cyclopropyl analog **16** was shown to be potent. However, the utilization of an aromatic substituent bearing a hydrogen bond acceptor at the *ortho* position proved to be advantageous; anisyl derivative **20**, in particular, showing high potency.

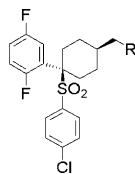
Sulfonamides were also well tolerated with azetidine derivative **24** being optimal in this series.

Whilst many of the 4-substituted cyclohexyl analogs prepared showed excellent *in vitro* potency, their *in vivo* efficacy profiles were often compromised by poor metabolic stability prompting us to investigate the cause of this issue. It was postulated that the activated methylene *exo* to the cyclohexyl core may be a site of metabolism and this was confirmed in the case of methyl analog **25** with the isolation of acid **26** as a major metabolite following *in vitro* incubation with rat liver microsomes (Fig. 1).

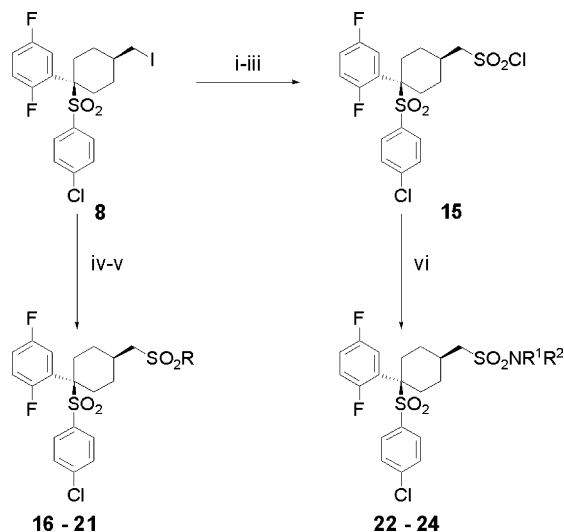
To address this potential route of metabolism, compounds lacking this labile methylene unit were next prepared. Initially, a wide range of functional groups were introduced directly linked to the cyclohexyl ring at the 4-position with the potency of a representative number summarized in Table 3. These compounds were available from the *syn* alcohol **3** (Scheme 1) or *syn* amine **7** (Scheme 3).

In most cases, the presence of a polar group directly attached to the cyclohexyl core was attended with inferior *in vitro* potency (note, in particular, the parent alcohol **3** and amine **7**). Acylation of **3** or **7** was also not productive, except in the case of carbamate **28** where potency was at best improved only to the level of unsubstituted **1**.

In marked contrast to acylated amino analogs, extending this work to a series of sulfonamides and sulfamates gave compounds with increased potency.

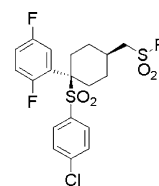
Table 1. In vitro γ -secretase inhibition of compounds 5–14

Compound	Preparation	R	IC ₅₀ (nM)
5	Scheme 1	OH	4.7 ± 1.7
6	Scheme 1	CO ₂ H	21.2 ± 1.5
7	Scheme 1	NH ₂	148 ± 28
8	Scheme 1	I	14.3 ± 7.2
9	5, 4-Nitrophenyl chloroformate, pyr, THF then MeNH ₂ /EtOH	OC(=O)NHMe	0.51 ± 0.15
10	7, Ac ₂ O, DMAP, Et ₃ N, DCM	NHAc	1.3 ± 0.2
11	6, DPPA, Et ₃ N, PhMe, 110 °C then NH ₃ /dioxan, rt	NHCONH ₂	1.0 ± 0.1
12	7, MeO ₂ CCl, Et ₃ N, DCM, 0 °C	NHCO ₂ Me	2.2 ± 0.1
13	7, catechol sulfate, THF then ethylamine/dioxan	NHSO ₂ NHET	1.7 ± 0.3
14	5, MsCl, Et ₃ N, DCM then succinimide, NaH, DMF, 80 °C		1.0 ± 0.4

**Scheme 2.** Reagents and conditions: (i) KSAc, DMF; (ii) NaOH, aq MeOH; (iii) KNO₃, SO₂Cl₂, MeCN; (iv) RSH, KOH, EtOH, reflux; (v) cat. RuO₂, NaIO₄, EtOAc/H₂O; (vi) R₁R₂NH, THF.

In the series of sulfonamides, alkyl substituents were well tolerated with *n*-propyl (**31**) and trifluoromethyl (**32**) substituents preferred. A series of aryl sulfonamides was also prepared and whilst the observation noted earlier of the favorable effect of introduction of *ortho*, electronegative substituents appeared not to translate to this series, several potent heterocyclic analogs (e.g., **33**) could be prepared. With some parallel to the earlier series of sulfamides (Table 2), this functional group was further utilized to give potent compounds with the azetidene derivative **35** giving extremely high levels of activity.

On consideration of in vitro potency and ADME profiles of the potent inhibitors, the trifluoromethane sulfonamide **32** was selected for further work. Compound **32** inhibited the generation of A β (40) and A β (42) with

Table 2. In vitro γ -secretase inhibition of compounds 16–24

Compound	R	IC ₅₀ (nM)
16	^c Pr	1.4 ± 0.3
17	CF ₃	21.5 ± 5
18	Ph	8.8 ± 1.7
19		0.65 ± 0.1
20		0.51 ± 0.02
21		2.1 ± 0.2
22	NH ₂	4.2 ± 1.4
23	NMe ₂	1.1 ± 0.3
24		0.36 ± 0.02

very similar IC₅₀s¹⁸ and inhibited the cleavage of Notch receptor¹⁹ at concentrations similar to those required for inhibition of APP processing,²⁰ consistent with the model that γ -secretase is required for both of these processes.



Figure 1. In vitro metabolism of methyl analog **25**.

This compound was profiled in vivo in the APP-YAC mouse model²¹ and demonstrated a robust decrease of DEA-extractable brain A β (40) levels^{22,23} 4 h after oral dosing with a minimum effective dose of 1 mg/kg (Fig. 2). A time-course study at 10 mg/kg showed excellent duration of action with a significant lowering of brain A β (40) levels throughout a 24 h period.

In summary, we have developed a series of 4-substituted cyclohexyl sulfones which inhibit γ -secretase in vitro in the low to sub-nanomolar range. This high level of in vitro potency could be coupled with good pharmacokinetics to produce potent, orally active γ -secretase

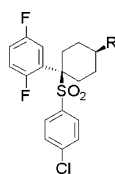
Scheme 3. Preparation of amine **27**. Reagents and conditions: (i) MsCl, Et₃N, DCM, -50 °C; (ii) NaN₃, DMF, 90 °C; (iii) PPh₃, aq THF.

inhibitors, such as **32**, which was shown to have excellent efficacy in reducing central A β (40) levels in APP-YAC mice with a minimum effective dose of 1 mg/kg p.o.

Acknowledgments

The authors thank Beth Oxley, Ian Gowers, and Robert Newman for assistance in screening.

Table 3. In vitro γ -secretase inhibition of compounds **27–35**



Compound	Preparation	R	IC ₅₀ (nM)
3	Scheme 1	OH	10.7 ± 1.7
28	3 , ClSO ₂ NCO, THF, 0 °C	OCONH ₂	1.9 ± 0.5
27	Scheme 3	NH ₂	510 ± 10
29	27 , Ac ₂ O, Et ₃ N, DCM	NHAc	35 ± 3
30	27 , MsCl, Et ₃ N, DCM	NHSO ₂ Me	4.2 ± 1.2
31	27 , PrSO ₂ Cl, Et ₃ N, DCM	NHSO ₂ Pr	0.82 ± 0.22
32	27 , Tf ₂ O, Et ₃ N, DCM	NHSO ₂ CF ₃	0.65 ± 0.13
33	27 , ArSO ₂ Cl, Et ₃ N, DCM		0.25 ± 0.07
34	27 , ArSO ₂ Cl, Et ₃ N, DCM		0.66 ± 0.01
35	27 , [reagent from azetidine/SO ₂ Cl ₂], Hunig's base, MeCN		0.15 ± 0.07

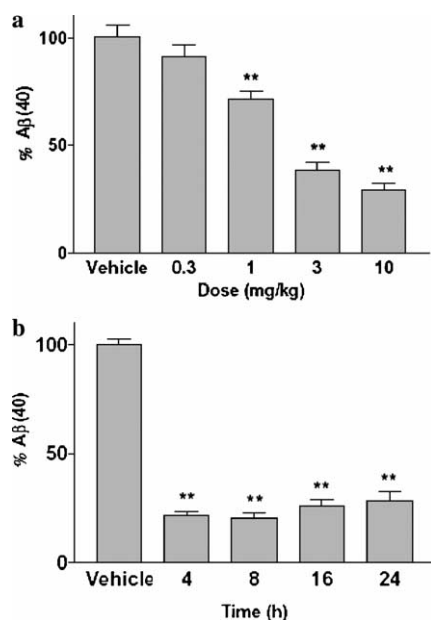


Figure 2. Effects of **32** on DEA-soluble brain Aβ(40) in APP-YAC mice. (a) Dose–response following p.o. dosing of **32** with analysis 4 h post-dose. (b) Time-course following 10 mg/kg p.o. dose. $n = 5$ per group. Compound was dosed as a suspension in 0.5% methocel solution.

References and notes

- Morgan, C.; Colombres, M.; Nunez, M. T.; Inestrosa, N. C. *Prog. Neurobiol.* **2004**, *74*, 323.
- Tanzi, R. E.; Bertram, L. *Cell* **2005**, *120*, 545.
- Bird, T. D. N. *Eng. J. Med.* **2005**, *352*, 862.
- Hardy, J. A.; Higgins, G. A. *Science* **1992**, *256*, 184.
- Russo, C.; Venezia, V.; Repetto, E.; Nizzari, M.; Violani, E.; Carlo, P.; Schettini, G. *Brain Res. Rev.* **2005**, *48*, 257.
- Harrison, T.; Churcher, I.; Beher, D. *Curr. Opin. Drug Discovery Dev.* **2004**, *7*, 709.
- Churcher, I.; Beher, D. *Curr. Pharm. Des.* **2005**, *11*, 3363.
- Pollack, S. J.; Lewis, H. *Curr. Opin. Investig. Drugs* **2005**, *6*, 35.
- van Es, J. H.; van Gijn, M. E.; Riccio, O.; van den Born, M.; Vooijs, M.; Begthel, H.; Cozijnsen, M.; Robine, S.;

- Winton, D. J.; Radtke, F.; Clevers, H. *Nature* **2005**, *435*, 959.
- Weng, A. P.; Lau, A. *Future Oncol.* **2005**, *1*, 511.
- Clarke, E. E.; Shearman, M. S. *J. Neurosci. Methods* **2000**, *102*, 61.
- Teall, M.; Oakley, P.; Harrison, T.; Shaw, D.; Kay, E.; Elliott, J.; Gerhard, U.; Castro, J. L.; Shearman, M.; Ball, R. G.; Tsou, N. N. *Bioorg. Med. Chem. Lett.* **2005**, *15*, 2685.
- Churcher, I.; Dinnell, K.; Harrison, T.; Kerrad, S.; Nadin, A. J.; Oakley, P. J.; Shaw, D. E.; Teall, M. R.; Williams, B. J.; Williams, S. Preparation of 1-phenyl-1-(arylsulfonyl)cyclohexanes for treatment of Alzheimer's disease. WO2003018543.
- Castro Pineiro, J. L.; Churcher, I.; Dinnell, K.; Harrison, T.; Kerrad, S.; Nadin, A. J.; Oakley, P. J.; Owens, A. P.; Shaw, D. E.; Teall, M. R.; Williams, B. J.; Williams, S. Preparation of aryl sulfones which modulate the action of gamma secretase. WO2002081435.
- Harrison, T.; Oakley, P. J.; Teall, M. R. Preparation of arylsulfones as modulators of gamma secretase. WO2002081433.
- Wong, M. K.; Chung, N. W.; He, L.; Wang, X. C.; Yan, Z.; Tang, Y. C.; Yang, D. *J. Org. Chem.* **2003**, *68*, 6321.
- St. Denis, Y.; Levesque, S.; Bachand, B.; Edmunds, J. J.; Leblond, L.; Preville, P.; Tarazi, M.; Winocour, P. D.; Siddiqui, M. A. *Bioorg. Med. Chem. Lett.* **2002**, *12*, 1181.
- Beher, D.; Clarke, E. E.; Wrigley, J. D. J.; Martin, A. C. L.; Nadin, A.; Churcher, I.; Shearman, M. S. *J. Biol. Chem.* **2004**, *279*, 43419.
- Kadesch, T. *Curr. Opin. Gen. Dev.* **2004**, *14*, 506.
- Lewis, H. D.; Perez Revuelta, B. I. P.; Nadin, A.; Neduvellil, J. G.; Harrison, T.; Pollack, S. J.; Shearman, M. S. *Biochemistry* **2003**, *42*, 7580.
- Lamb, B. T.; Sisodia, S. S.; Lawler, A. M.; Slunt, H. H.; Kitt, C. A.; Kearns, W. G.; Pearson, P. L.; Price, D. L.; Gearhart, J. D. *Nat. Genet.* **1993**, *5*, 22.
- Best, J. D.; Jay, M. T.; Otu, F.; Ma, J.; Nadin, A.; Ellis, S.; Lewis, H. D.; Pattison, C.; Reilly, M.; Harrison, T.; Shearman, M. S.; Williamson, T. L.; Atack, J. R. *J. Pharmacol. Exp. Ther.* **2005**, *313*, 902.
- Savage, M. J.; Trusko, S. P.; Howland, D. S.; Pinsker, L. R.; Mistretta, S.; Reaume, A. G.; Greenberg, B. D.; Siman, R.; Scott, R. W. *J. Neurosci.* **1998**, *18*, 1743.